Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry

journal homepage: www.elsevier.com/locate/bmc

Synthesis and biological evaluation of 3-substituted 5-benzylidene-1methyl-2-thiohydantoins as potent NADPH oxidase (NOX) inhibitors

Yun Soo Bae^{a,†}, Sun Choi^{b,†}, Jung Jae Park^c, Jung Hee Joo^a, Minghua Cui^b, Hyunsung Cho^c, Won Jae Lee^d, Sang Hyup Lee^{c,*}

^a Department of Life Science, College of Natural Sciences, Ewha Womans University, Seoul 03760, Republic of Korea

^b College of Pharmacy and Graduate School of Pharmaceutical Sciences, Ewha Womans University, Seoul 03760, Republic of Korea

^c College of Pharmacy and Innovative Drug Center, Duksung Women's University, Seoul 01369, Republic of Korea

^d Department of Life Science, Seoul National University, Seoul 08826, Republic of Korea

ARTICLE INFO

Article history: Received 2 June 2016 Revised 28 June 2016 Accepted 29 June 2016 Available online 30 June 2016

Keywords: Thiohydantoin Benzylidene Aluminum chloride NADPH oxidase (NOX)

ABSTRACT

We report the synthesis of novel 3-substituted 5-benzylidene-1-methyl-2-thiohydantoins **3**, and their biological evaluation using NADPH oxidase (NOX) 1 and 4. Based on structural and pharmacophore analyses of known inhibitors such as hydroxypyrazole **2**, we envisioned interesting 2-thiohydantoin compounds, 3-substituted 5-benzylidene-1-methyl-2-thiohydantoins **3** that would be expected to well match the structural features in **2**. Efficient synthesis of eighteen target compounds **3** were achieved through the synthetic pathway of $\mathbf{4} \rightarrow \mathbf{11} \rightarrow \mathbf{3}$, established after consideration of several plausible synthetic pathways. The inhibitory activities of compounds **3** against NOX 1 and 4 were measured, with some of the target compounds showing similar or higher activities compared with reference **2**; in particular, compounds **3bz**, **3cz**, and **3ez** were found to be promising inhibitors of both NOX 1 and 4 with modest isozyme selectivities, which highlights the significance of the 2-thiohydantoin substructure for inhibition of NOX 1 and 4. This marks the first time these compounds have been applied to the inhibition of NOX enzymes.

© 2016 Elsevier Ltd. All rights reserved.

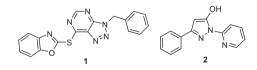
1. Introduction

The 5-benzylidene-2-thiohydantoin compounds have been found to display a range of biological activities, such as antimycobacterial,¹ antiviral,² anticonvulsant,³ and antitumor properties.⁴ Among various thiohydantoin compounds we were interested in 3substituted 5-benzylidene-1-methyl-2-thiohydantoins as potential inhibitors of NADPH oxidase (NOX) 1 and 4. NOX enzymes are known to generate reactive oxygen species (ROS) that play a significant role in many kinds of pathogenesis such as inflammation and vascular and fibrotic disorders.⁵ NOX 1 seems to play a significant role in CNS diseases.³ NOX 4 appears to be the most abundant isoform in endothelial and vascular smooth muscle cells,^{6,7} and a strong correlation between expression of NOX 4 and total NOX activities in human coronary arteries was observed.⁸ Various natural and synthetic compounds display NOX inhibitory activities,⁹ and the general results were reviewed in detail by Borbely et. al.¹⁰ A triazolo[4,5-d]pyrimidine derivative **1** (VAS2870)^{9,11} was

E-mail address: sanghyup@duksung.ac.kr (S.H. Lee).

[†] These authors contributed equally to this paper.

found to inhibit oxLDL-induced superoxide release from human endothelial cells.¹² More significantly, a hydroxypyrazole derivative 2^{13-16} was recently disclosed to have potent activities against NOX enzymes.¹⁵



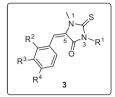
Although the results thus far seem reasonably good and a substantial amount of information has been accumulated, there are still no potent and selective inhibitors of NOX 1 and 4. Therefore, with the aim to develop promising inhibitors against NOX 1 and 4, we performed preliminary studies through analysis of the structural features of known inhibitors such as **1** and **2**, library searches, and molecular modeling, which eventually guided us to focus on the thiohydantoin substructure. Here, we present studies on the synthesis of 3-substituted 5-benzylidene-1-methyl-2-thiohydantoins





^{*} Corresponding author. Tel.: +82 2 901 8393.

3 and their inhibitory activities against NOX 1 and 4, which is the first application of these target compounds to the inhibition of NOX enzymes.



2. Results and discussion

2.1. Synthesis of 2-thiohydantoins 3

We began a program to develop potent inhibitors of NOX 1 and 4 starting from known analogues such as hydroxypyrazole 2. Based on the structural analysis of 2 and related derivatives (data not shown), we obtained information on the structural features and important pharmacophores. For example, a consecutive arrangement of rigid ring structures, a hydrophilic region including hydrogen bond acceptors (P1), hydrophobic side chains (S1) near N(3), and extra hydrophobic pockets (S2) near C(5) may be important for activity (Fig. 1). Through these studies, we envisioned a structurally similar moiety, thiohydantoin, and consequently, suggested novel 3-substituted 5-benzylidene-1-methyl-2-thiohydantoins 3 as NOX inhibitors. When we compared thiohydantoins 3 with the reference 2, we recognized that the thiohydantoin core, ring substituent R^1 at N(3), and benzylidene group at C(5) in **3** would correlate well with the hydroxypyrazole, pyridinyl, and phenyl groups in 2, respectively. In particular, we tried to align the thiohydantoin ring to best overlap with the hydroxypyrazole ring according to the pharmacophore analysis. Thus, we introduced ring substituents such as cycloalkyl or aryl group at N(3) and incorporated the benzylidene group at C(5) for a better match with the reference compound, to finally afford the target thiohydantoins **3**. Although the benzylidene group appeared to be slightly different from the phenyl group in 2, we believed that a large group could be accommodated in this region (S2) according to the previous studies.¹⁵

In order to establish an efficient synthetic method of target compounds **3**, we evaluated several retrosynthetic pathways, as shown in Scheme 1. First, a starting material sarcosine (**4**) could be transformed to thiourea **5**,¹⁷ which could be further converted to 2-thiohydantoin **6**.¹⁷ Alternatively, compound **6** could be prepared from hydantoin **7**. Compound **6** needs to undergo both C (5)-functionalization and N(3)-alkylation to yield target compounds **3**. Thus, C(5)-functionalization using the corresponding benzaldehydes **8** would give 5-benzylidene-2-thiohydantoins **9**, and N(3)-alkylation using alkyl halide (R¹X) would lead to the generation of compounds **3**. Alternatively, N(3)-alkylation of **6** would give the key intermediate, 3-substituted thiohydantoins **11**, and further C(5)-functionalization would provide **3**. Additionally, the key intermediates **11** could also be synthesized from coupling of **4** and alkyl isothiocyanates **10**.

In order to investigate the feasibility of these synthetic pathways, we briefly checked their practical applicability by carrying out several model reactions. By treating **4** and ammonium thiocyanate (NH₄SCN) as previously described,¹⁷ we obtained the thiohydantoin **6** in relatively low yield (30–40%). We then applied another method in which hydantoin **7** was treated with phosphorous pentasulfide (P_2S_5)¹⁸ to give compound **6** in moderate yield (~60%). Next, we attempted N(3)-alkylation to convert **6** to **11**

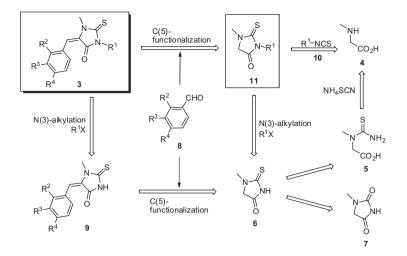
S2 R^{2} R^{2} R^{4} R^{4} R^{1} R^{1} R^{1} R^{2} R^{2}

Figure 1. Pharmacophores of 2 and comparison with 3.

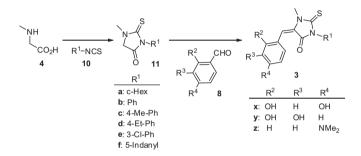
using an analogous procedure;¹⁹ however, the N(3)-alkylation of **6** with alkyl halide (e.g., cyclohexyl bromide) did not provide good results, as several side products were generated along with the desired product. We found that the reaction proceeded with the formation of several polar spots on TLC; the side product(s) might be generated by S-alkylation, which derives some support from previous work.^{20,21} If this is the case, then N(3)-alkylation could be a problematic step in the whole pathway. In addition, we believed that, in order to avoid interference of N(3)H in C(5)-functionalization, it would be more advantageous to perform the N(3)-alkylation prior to C(5)-functionalization rather than the reverse order of transformations.

Considering the efficiency of each pathway and the preliminary results of the model reactions, we adopted the pathway of $4 \rightarrow 11 \rightarrow 3$. Therefore, for the synthesis of kev intermediates 11. we employed the previous procedures^{22,23} with minor modifications. As shown in Scheme 2, sarcosine (4) was treated with alkyl isothiocvanates 10 to provide 3-substituted 1-methyl-2-thiohydantoins 11 in modest yields. In these reactions, cyclohexyl (a), phenyl (**b**), 4-methylphenyl (**c**), 4-ethylphenyl (**d**), 3-chlorophenyl (e), and indan-5-yl group (f) were used as a set of \mathbb{R}^1 groups at $\mathbb{N}(3)$. For the synthesis of indan-5-yl isothiocyanate (10f), N-indan-5yl-O-phenyl thiocarbamate, prepared from 5-aminoindane and phenyl chlorothionoformate, was treated with trichlorosilane and triethylamine to afford **10f** in 50% yield (for two steps).²⁴ The reactions to synthesize 2-thiohydantoins 11 generally provided good results (more than 90% yield), except in the cases of the cyclohexyl (11a, 66% yield) and 3-chlorophenyl groups (11e, 74% yield), probably due to their electronic and steric effects on the reactivity of nitrogen in isothiocyanate. As a result, the six key intermediates, 11a,²⁵ 11b,^{22,26} 11c,²⁷ 11d,²⁸ 11e, and 11f²² were efficiently synthesized.

We then investigated several methods to perform the condensations of key intermediates 11 with aldehydes 8 to provide the products 3. Here, three aromatic aldehydes, 2,4dihydroxybenzaldehyde (8x), 2,3-dihydroxybenzaldehyde (8y), and 4-(*N*.*N*-dimethylamino)benzaldehyde (**8z**) were employed. At first, the method using melt sodium acetate in acetic acid¹⁹ was applied to our substrates, which gave unsuccessful results probably due to the different structures in the thiohydantoins and aldehydes. Next, microwave-assisted synthesis²⁹ was found to give moderate results with a limited range of substrates. Furthermore, the condensation reactions were conducted by simply using a secondary amine such as piperidine or pyrrolidine; however, we were only partially successful in obtaining the product with some unhindered aldehydes in moderate to low yield (less than 60% yield). Acid catalysts have also been used for similar reactions with different substrates (not thiohydantoins), and furthermore, Greg et al. achieved improved results by employing Lewis acid catalysts^{23,30} compared to the same reactions without acid catalysts.



Scheme 1. Retrosynthetic analysis of 2-thiohydantoins 3.



Scheme 2. Synthesis of 2-thiohydantoins 3.

Therefore, we investigated reaction conditions using our substrates and acid catalysts in the presence of piperidine at 25-80 °C. First, reactions of **11a** and **8x** using acid catalysts such as SnCl₄, TiCl₄, AlCl₃, and MgBr₂ to afford **3ax** were examined. When a catalytic amount (0.1-0.5 equiv) of SnCl₄ or TiCl₄ was employed, the starting materials **11a** and **8x** gradually disappeared; however, although several side products were generated, product **3ax** was not formed. When MgBr₂ was used, the results of the reaction were inconclusive; a small amount of product formed but subsequently disappeared. When AlCl₃ was applied, the product **3ax** was formed in good to modest yields (\sim 60%) with an appropriate reaction time and temperature. Considering the varying results with substrate and reaction conditions, we believed that polar hydroxy groups might affect the function of the acid catalyst, presumably by lowering its activity. Thus, the reaction was tested using 8z, that does not contain a hydroxy group, to afford **3az** in an improved yields (~85%) compared to **3ax** (~60%).

Having chosen to use AlCl₃, we synthesized the target compounds **3** with minor modifications to the reaction conditions and work-up procedures. Thus, AlCl₃ (0.1 equiv) in the presence of piperidine (1.5 equiv) at 65–80 °C was used for the condensation of **11** and **8** to synthesize the target compounds **3**. In general, the reaction mixtures were directly applied to a column chromatography after completion of the reaction. However, in some cases (e.g., **3ey** and **3ez**) the reaction mixtures were subjected to workup procedures to give the crude products that were subsequently applied to column chromatography to afford **3**. Notably, compounds **3** contain an exocyclic C=C bond and could therefore exist as *Z/E*-isomers; however, information on the isomeric mixture of 5-benzylidene-2-thiohydantoins has not been revealed except in a few cases.^{29,31} In our experiments, we observed the formation of a single isomer in some cases and a mixture of isomers in the other cases based on ¹H and ¹³C NMR analyses. However, the identification of isomer structures and their separation would require further intensive work, which was not within the scope of our present studies. Consequently, we have achieved the efficient synthesis of eighteen 2-thiohydantoin compounds **3**. Among these, compound **3fz** was previously reported²³ and compound **3bz** was simply introduced without any characterization.³ Compounds **3ax**, **3az**, **3by**, **3fx**, and **3fy** were known as members of chemical libraries,^{28,32} but no information on their preparation, isomer ratio, and spectral data were disclosed. To the best of our knowledge, the other eleven compounds have not been previously identified with appropriate characterization.

2.2. Evaluation of NOX 1 and 4 inhibitory activities and SAR studies

We evaluated the NOX 1 and 4 inhibitory activities of the synthesized compounds **3** along with hydroxypyrazole derivative **2** as a reference. This marks the first time these compounds have been applied to the inhibition of NOX enzymes. To obtain specific inhibition of NOX 1 and 4 enzymes, we established transgenic fly lines overexpressing human NOX 1 or human NOX 4 in a Drosophila DUOX knockdown condition according to our previous procedure.³³ Transgenic flies were homogenated with ice-cold PBS containing protease inhibitors, and the membranes enriched human NOX 1 or 4 were harvested. The membranes with human NOX 1 or 4 served as ROS production monitoring by lucigenin chemiluminescence in the absence or presence of chemical compounds.³⁴ The results of inhibitory activities for all of the thiohydantoins against NOX 1 and 4 were shown in Table 1. We first evaluated **3ax**, **3ay**, and **3az** (**3a** series) containing a cyclohexyl group at N(3) as an aliphatic chain (R^1) and three benzylidene groups at C(5), respectively. For NOX 1, compound 3ay that contains a 2,3dihydroxybenzylidene group exhibited higher activity than 3ax and **3az**, but, lower activity than reference **2**. Inhibitory activities of the compounds in the **3a** series also displayed similar trends for NOX 4. Next, we evaluated **3bx**. **3bv**. **3bz** (**3b** series). **3cx**. **3cv**. 3cz (3c series), and 3dx, 3dy, 3dz (3d series) containing phenyl, 4-methylphenyl, and 4-ethylphenyl groups at N(3) as nonpolar aromatic groups (R¹), and the same three benzylidene groups at C(5), respectively. In the **3b** series, while compounds **3bx** and 3by displayed moderate to low activities for NOX 1 and 4, compound **3bz** with a 4-(*N*,*N*-dimethylamino)benzylidene group showed excellent activities that were similar or higher compared

Table 1Inhibitory activities of compounds 3 for NOX 1 and 4

Compound		Inhibitory activity (K_i , μ M)	
		NOX 1	NOX 4
3a series	3ax	26.0 ± 1.2	37.6 ± 0.76
	3ay	7.39 ± 3.6	6.35 ± 2.4
	3az	12.3 ± 19.3	35.6 ± 14
3b series	3bx	33.8 ± 4.7	27.7 ± 12
	3by	49.7 ± 4.4	24.0 ± 6.3
	3bz	1.95 ± 0.07	0.88 ± 0.01
3c series	3cx	27.1 ± 7.6	34.6 ± 4.4
	3cy	50.3 ± 33	27.0 ± 2.8
	3cz	1.02 ± 0.07	2.33 ± 0.05
3d series	3dx	8.40 ± 0.27	2.39 ± 0.02
	3dy	27.9 ± 6.1	26.4 ± 5.7
	3dz	4.39 ± 0.06	2.63 ± 0.13
3e series	3ex	34.5 ± 3.3	33.8 ± 8.1
	3ey	7.65 ± 1.4	9.31 ± 0.10
	3ez	0.35 ± 0.04	0.84 ± 0.06
3f series	3fx	6.58 ± 0.33	6.83 ± 8.6
	3fy	6.63 ± 6.0	2.36 ± 1.9
	3fz	8.04 ± 2.5	13.1 ± 6.3
2 ^a		1.45 ± 0.15^{a}	$3.12 \pm 0.52^{\circ}$

^a Ref. 15.

to reference 2 [\sim 3.5 fold (NOX 4)]. Trends in the 3c series were similar to the **3b** series, but compound **3cz** exhibited higher activities in both NOX 1 and 4 than reference 2 [~1.4 fold (NOX 1) and \sim 1.3 fold (NOX 4)]. A similar trend was observed in the **3d** series. Compound **3dz** showed similar or higher activities compared to reference 2 and notably, 3dx containing a 2,4-dihydroxybenzylidene group also displayed higher activity than 2 against NOX 4. We then evaluated **3ex**, **3ey**, and **3ez** (**3e** series) containing 3-Clphenyl group at N(3) as a slightly different group to see the effect of bulky meta-substituents. Surprisingly, among the compounds tested, compound **3ez** displayed the highest activities that were higher than reference 2 against both NOX 1 and 4 [~4.1 fold (NOX 1) and \sim 3.7 fold (NOX 4)]. Finally, we evaluated **3fx**, **3fy**, and 3fz (3f series) containing indan-5-yl group at N(3) to assess the effect of an additional fused ring. In general, these compounds showed good to modest activities, and compound 3fy displayed higher activities than 2 for NOX 4. As for isozyme selectivities between NOX 1 and 4, compounds 3cz and 3ez showed modest selectivities for NOX 1 (2.3 and 2.4 fold, respectively), and compounds 3bz, 3dx and 3fy showed modest selectivities for NOX 4 (2.2, 3.5 and 2.8 fold, respectively).

From the inhibition results, it was evident that the three types of substituents at C(5) strikingly affected the activities. While compounds containing a dihydroxybenzylidene group (\mathbf{x} or \mathbf{y}) at C(5) gave low to modest activities, compounds containing a 4-(N,N-dimethylamino)benzylidene group (z) provided the highest activities in most of the series. Considering these results, a nonpolar hydrogen bond acceptor (e.g., *N*,*N*-dimethylamino group) at C(5) was highly preferable rather than a polar hydrogen bond donor (e.g., dihydroxy group). However, the six types of substituents at N(3) did not seem to significantly affect the activities, and the 3-chlorophenyl (3e series) and indan-5-yl (3f series) groups gave slightly better results than the other series. Consequently, it was found that compounds 3bz, 3cz and 3ez displayed the highest level of activities against both NOX 1 and 4, similar or higher compared to reference 2, along with modest isozyme selectivity. Thus, we have successfully developed improved, promising inhibitors of NOX 1 and 4 using a combination of a 4-(N,N-dimethylamino)benzylidene group at C (5) and a phenyl, 4-methylphenyl, or 3-chlorophenyl group at N (3), respectively.

3. Conclusion

In this study, we synthesized novel 3-substituted 5-benzylidene-1-methyl-2-thiohydantoins **3** and investigated their inhibitory activities against NOX 1 and 4. Compounds 3 were designed based on an analysis of the structures of existing inhibitors such as reference 2 and information on their pharmacophores, and thus, were expected to match the structural features found in 2. We achieved the efficient synthesis of target compounds **3** through the synthetic pathway of $4 \rightarrow 11 \rightarrow 3$, to provide eighteen 2-thiohydantoin compounds 3. We then measured the inhibitory activities of compounds 3 against NOX 1 and 4, and found that compounds 3bz, 3cz and 3ez showed the highest levels of activity with modest isozyme selectivities that were similar or higher compared to reference 2, which suggests they are promising compounds as inhibitors of NOX 1 and/or 4. This marks the first time these compounds have been applied to the inhibition of NOX enzymes. Thus, the targeted 2-thiohydantoin moiety was demonstrated to be a meaningful substructure for the inhibition of NOX 1 and 4.

4. Experimental section

4.1. General information

¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded on a Bruker DRX 300 spectrometer. Mass spectra were obtained by the CI, EI or FAB ionization method. FT-IR spectra were run on a Perkin-Elmer Spectrum GX spectrometer. Melting points were determined in open capillary tubes using a Buchi B-545 melting point apparatus and were uncorrected. Thin layer chromatography was performed with general purpose silica gel plates (20×20 cm; Aldrich No. Z12272-6). Deionized water was obtained with a Milli-Q ($18 M\Omega$ s) water system (Millipore). The solvents and reactants were of the highest commercial grade available and were used without further purification unless noted.

4.2. General procedure^{22,23} for preparation of 3-substituted 1methyl-2-thiohydantoins 11

To a stirred solution of sarcosine (**4**, 180 mg, 2.0 mmol) in anhydrous ethanol (6 mL) was added the appropriate isothiocyanate (**10**, 2.0 mmol). The reaction mixture was refluxed for 5 h and then was allowed to cool to room temperature. The solvent was removed in vacuo, and the resulting residue was purified by column chromatography (EtOAc/hexanes) to afford the title compound.

4.2.1. 3-Cyclohexyl-1-methyl-2-thiohydantoin (11a)

Use of cyclohexyl isothiocyanate (**10a**) and the general procedure (85 °C, 6 h) afforded the title compound as a yellow solid: yield, 281 mg (66%); Mp 160–162 °C; R_f 0.36 (1:2 EtOAc/hexanes); ¹H NMR (300 MHz; DMSO- d_6) δ : 0.93–1.35 (m, 3H, *c*-Hx), 1.50–1.65 (m, 3H, *c*-Hx), 1.68–1.84 (m, 2H, *c*-Hx), 1.98–2.20 (m, 2H, *c*-Hx), 3.18 (s, 3H, N(1)CH₃), 4.17 (s, 2H, C(5)CH₂); ¹³C NMR (75 MHz; DMSO- d_6) δ : 24.8 (*c*-Hx), 25.5 (*c*-Hx), 28.2 (*c*-Hx), 33.9 (N(1)CH₃), 53.5 (C(5)), 55.0 (N(3)CH), 171.3 (C=O), 182.1 (C=S). MS m/z 212 [M]⁺. HRMS (EI+) calcd for C₁₀H₁₆N₂OS [M]⁺: 212.0983; found 212.0982.

4.2.2. 3-Phenyl-1-methyl-2-thiohydantoin (11b)²²

Use of phenyl isothiocyanate (**10b**) and the general procedure (85 °C, 2 h) afforded the title compound as a yellow solid: yield, 375 mg (91%); Mp 163–165 °C; R_f 0.25 (1:2 EtOAc/hexanes); ¹H NMR (300 MHz; CDCl₃) δ : 3.34 (s, 3H, N(1)CH₃), 4.16 (s, 2H, C(5)

H₂), 7.18–7.26 (m, 2H, Ph), 7.34–7.48 (m, 3H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ: 34.7 (N(1)CH₃), 54.7 (C(5)), 128.7, 129.5, 129.6, 133.7 (Ph), 170.0 (C=O), 183.6 (C=S). MS *m*/*z* 206 [M]⁺. HRMS (EI+) calcd for C₁₀H₁₀N₂OS [M]⁺: 206.0513; found 206.0515.

4.2.3. 3-(4-Methylphenyl)-1-methyl-2-thiohydantoin (11c)²⁷

Use of 4-methylphenyl isothiocyanate (**10c**) and the general procedure (85 °C, 2 h) with recrystallization (EtOAc/hexanes) instead of column chromatography afforded the title compound as a yellow solid: yield, 405 mg (92%); Mp 147–149 °C; R_f 0.29 (1:2 EtOAc/hexanes); ¹H NMR (300 MHz; CDCl₃) δ : 2.32 (s, 3H, PhCH₃), 3.31 (s, 3H, N(1)CH₃), 4.13 (S, 2H, C(5)H₂), 7.10 (d, J = 6.3 Hz, 2H, Ph), 7.22 (d, J = 6.3 Hz, 2H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 21.4 (PhCH₃), 34.4 (N(1)CH₃), 54.5 (C(5)), 128.1, 130.0, 130.8, 139.5 (Ph), 170.0 (C=O), 183.4 (C=S). MS m/z 220 [M]⁺. HRMS (El+) calcd for C₁₁H₁₂N₂OS [M]⁺: 220.0670; found 220.0668.

4.2.4. 3-(4-Ethylphenyl)-1-methyl-2-thiohydantoin (11d)

Use of 4-ethylphenyl isothiocyanate (**10d**) and the general procedure (85 °C, 1 h) afforded the title compound as a yellow solid: yield, 422 mg (90%); Mp 115–117 °C; R_f 0.16 (1:2 EtOAc/hexanes); ¹H NMR (300 MHz; CDCl₃) δ : 1.28 (t, J = 7.6 Hz, 3H, PhCH₂CH₃), 2.71 (q, J = 7.6 Hz, 2H, PhCH₂), 3.42 (s, 3H, N(1)CH₃), 4.23 (s, 2H, C(5)H₂), 7.22 (d, J = 8.2 Hz, 2H, Ph), 7.34 (d, J = 8.2 Hz, 2H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 15.3 (PhCH₂CH₃), 28.8 (PhCH₂), 34.5 (N(1)CH₃), 54.4 (C(5)), 128.2, 128.9, 131.0, 145.6 (Ph), 170.0 (C=O), 183.6 (C=S). MS m/z 234 [M]⁺. HRMS (EI+) calcd for C₁₂H₁₄N₂OS [M]⁺: 234.0826; found 234.0825.

4.2.5. 3-(3-Chlorophenyl)-1-methyl-2-thiohydantoin (11e)

Use of 3-chlorophenyl isothiocyanate (**10e**) and the general procedure (85 °C, 3 h) with recrystallization (EtOAc/hexanes) instead of column chromatography afforded the title compound as a yellow solid: yield, 356 mg (74%); Mp 192–194 °C; R_f 0.16 (1:2 EtOAc/hexanes); ¹H NMR (300 MHz; CDCl₃) δ : 3.41 (s, 3H, N(1) CH₃), 4.24 (s, 2H, C(5)H₂), 7.47–7.21 (m, 4H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 34.5 (N(1)CH₃), 54.4 (C(5)), 126.8, 128.8, 129.6, 130.2, 134.4, 134.7 (Ph), 169.5 (C=O), 182.6 (C=S). MS *m*/*z* 240 [M]⁺. HRMS (EI+) calcd for C₁₀H₉ClN₂OS [M]⁺: 240.0124; found 240.0122.

4.2.6. 3-(Indan-5-yl)-1-methyl-2-thiohydantoin (11f)²²

Use of indan-5-yl isothiocyanate (**10f**) and the general procedure (85 °C, 1 h) afforded the title compound as a yellow solid: yield, 473 mg (96%); Mp 158–160 °C; R_f 0.14 (1:2 EtOAc/hexanes); ¹H NMR (300 MHz; CDCl₃) δ : 2.07–2.28 (m, 2H, CH₂-indanyl), 2.87–3.01 (m, 4H, CH₂-indanyl), 3.43 (s, 3H, N(1)CH₃), 4.23 (s, 2H, C(5)H₂), 7.04 (dd, *J* = 7.8, 1.2 Hz, 1H, Ph-indanyl), 7.12 (s, 1H, Ph-indanyl), 7.34 (d, *J* = 7.8 Hz, 1H, Ph-indanyl); ¹³C NMR (75 MHz; CDCl₃) δ : 25.8, 33.1, 33.2 (CH₂-indanyl), 34.7 (N(1)CH₃), 54.7 (C(5)), 124.6, 125.3, 126.5, 131.6, 145.9, 146.2 (Ph-indanyl), 170.4 (C=O), 184.0 (C=S). MS *m/z* 246 [M]^{*}. HRMS (El+) calcd for C₁₃H₁₄N₂OS [M]^{*}: 246.0826; found 246.0828.

4.3. General procedure²³ for 3-substituted 5-benzylidene-1-methyl-2-thiohydantoins 3

To a stirred solution of thiohydantoin (**11**, 0.20 mmol), aldehyde (**8**, 0.24 mmol) and piperidine (30μ L, 0.30 mmol) in dioxane (0.6 mL) was added aluminum chloride (2.7 mg, 0.020 mmol). The stirred reaction mixture was heated to 65–80 °C until completion of the reaction. In cases where the reaction was incomplete, more aluminum chloride (0.1–0.2 equiv) was added along with the corresponding amount of piperidine in the same ratio, and the reaction mixture was refluxed for additional time. After cooling

to room temperature, the reaction mixture was directly subjected to column chromatography (gradient elution with appropriate EtOAc/hexanes mixtures) to give the title compounds **3**. In the cases of **3ey** and **3ez**, the reaction mixture was worked-up prior to column chromatography. The reaction mixture was evaporated to approximately half of the original volume. H₂O (50 mL) was slowly added, and the resulting mixture was extracted with EtOAc (2×50 mL). The combined organic layers were washed with saturated aqueous NaHCO₃ and brine (50 mL), dried (MgSO₄), and concentrated in vacuo to give an oily residue. Purification of the residue by column chromatography (gradient elution with appropriate EtOAc/hexanes mixtures) afforded the title compound.

4.3.1. 3-Cyclohexyl-5-(2,4-dihydroxybenzylidene)-1-methyl-2thiohydantoin (3ax)

Use of thiohydantoin **11a** and aldehyde **8x** in the general procedure (80 °C, 3 h) afforded the title compound as a red solid: yield, 43 mg (64%); Mp 205–207 °C; R_f 0.13 (1:2 EtOAc/hexanes); IR (KBr) 3434, 2090, 1637, 1384, 1096 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 1.10–1.35 (m, 3H, *c*-Hx), 1.56–1.70 (m, 3H, *c*-Hx), 1.75–1.87 (m, 2H, *c*-Hx), 2.15–2.29 (m, 2H, *c*-Hx), 3.53 (s, 3H, N(1)CH₃), 4.53–4.66 (m, 1H, N(3)CH), 6.28 (dd, *J* = 8.8, 2.2 Hz, 1H, Ph), 6.37 (d, *J* = 2.2 Hz, 1H, Ph), 6.97 (s, 1H, CH=C), 8.43 (d, *J* = 8.8 Hz, 1H, Ph), 10.10 (br s, 2H, OH); ¹³C NMR (75 MHz; DMSO- d_6) δ : 24.9 (*c*-Hx), 25.6 (*c*-Hx), 28.2 (*c*-Hx), 31.0 (N(1)CH₃), 54.8 (N(3)CH), 101.7, 107.0, 110.8, 116.9, 125.2, 132.1, 158.8, 161.4 (Ph, CH=C), 161.2 (C=O), 174.2 (C=S). MS *m/z* 332 [M]⁺. HRMS (EI+) calcd for C₁₇H₂₀N₂O₃S [M]⁺: 332.1194; found 332.1195.

4.3.2. 3-Cyclohexyl-5-(2,3-dihydroxybenzylidene)-1-methyl-2thiohydantoin (3ay)

Use of thiohydantoin **11a** and aldehyde **8y** in the general procedure (80 °C, 9 h) afforded the title compound as a red solid: yield, 46 mg (72%); Mp 95–97 °C; R_f 0.40 (1:2 EtOAc/hexanes); IR (KBr) 3433, 2930, 2854, 1635, 1471, 1402, 1372, 1283, 1188, 1146, 1099, 728, 530 cm⁻¹; ¹H NMR (300 MHz; CDCl₃) δ : 1.20–1.46 (m, 3H, *c*-Hx), 1.63–1.79 (m, 3H, *c*-Hx), 1.81–1.93 (m, 2H, *c*-Hx), 2.15–2.35 (m, 2H, *c*-Hx), 3.32, 3.66 (2s, 3H, N(1)CH₃), 4.56–4.84 (m, 1H, N(3)CH), 6.64, 6.84 (2s, 1H, CH=C), 6.69–6.83 (m, 1H, Ph), 6.87–7.08 (m, 2H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 25.2 (*c*-Hx), 26.1 (*c*-Hx), 28.8 (*c*-Hx), 31.7, 34.5 (N(1)CH₃), 56.4, 56.7 (N (3)CH), 109.7, 116.2, 117.8, 120.2, 120.3, 122.0, 122.4, 122.6, 124.8, 128.6, 130.8, 143.1, 143.3, 144.2, 148.7 (Ph, CH=C, one peak was not detected and believed to overlap with the observed peaks), 163.9, 165.4 (C=O), 176.1, 180.5 (C=S). MS *m*/*z* 332 [M]⁺. HRMS (EI +) calcd for C₁₇H₂₀N₂O₃S [M]⁺: 332.1194; found 332.1195.

4.3.3. 3-Cyclohexyl-5-(4-(*N*,*N*-dimethylamino)benzylidene)-1methyl-2-thiohydantoin (3az)

Use of thiohydantoin **11a** and aldehyde **8z** in the general procedure (65 °C, 6 h) afforded the title compound as a red solid: yield, 59 mg (86%); Mp 215–217 °C; R_f 0.57 (1:2 EtOAc/hexanes); IR (KBr) 2925, 1701, 1588, 1530, 1443, 1395, 1341, 1233, 1185, 1078, 950, 875, 807, 515 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 1.09–1.42 (m, 3H, *c*-Hx), 1.57–1.73 (m, 3H, *c*-Hx), 1.76–1.89 (m, 2H, *c*-Hx), 2.12–2.35 (m, 2H, *c*-Hx), 2.98, 3.01 (2s, 6H, N(CH₃)₂), 3.41, 3.56 (2s, 3H, N(1)CH₃), 4.41–4.67 (m, 1H, N(3)CH), 6.68–6.85 (m, 3H, Ph, CH=C), 7.38 (d, *J* = 9.0 Hz, 1H, Ph), 8.16 (d, *J* = 9.0 Hz, 1H, Ph); ¹³C NMR (75 MHz; DMSO- d_6) δ : 24.5, 24.9 (*c*-Hx), 25.5, 25.6 (*c*-Hx), 28.2, 28.3 (*c*-Hx), 31.1, 35.6 (N(1)CH₃), 54.8, 55.2 (N(3)CH), 111.2, 111.4, 116.8, 118.7, 119.9, 123.8, 124.6, 125.8, 132.0, 133.4, 150.6, 151.3 (Ph, CH=C), 161.1, 163.1 (C=O), 173.5, 178.6 (C=S). MS *m/z* 343 [M]⁺. HRMS (EI+) calcd for C₁₉H₂₅N₃OS [M]⁺: 343.1718; found 343.1718.

4.3.4. 3-Phenyl-5-(2,4-dihydroxybenzylidene)-1-methyl-2thiohydantoin (3bx)

Use of thiohydantoin **11b**, aldehyde **8x**, and additional AlCl₃ (0.020 mmol) and piperidine (0.30 mmol) in the general procedure (80 °C, 1 d and additionally, 10 h) afforded the title compound as a red solid: yield, 37 mg (57%); Mp 263–265 °C; R_f 0.25 (1:1 EtOAc/hexanes); IR (KBr) 3162, 2918, 2849, 2348, 2110, 1635, 1454, 1396, 1260, 1023, 873, 749, 538 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 3.61 (s, 3H, N(1)CH₃), 6.23 (dd, *J* = 8.8, 2.2 Hz, 1H, Ph), 6.37 (d, *J* = 2.2 Hz, 1H, Ph), 7.05 (s, 1H, CH=C), 7.28–7.63 (m, 5H, Ph), 8.41 (d, *J* = 8.8 Hz, 1H, Ph), 10.01, 10.27 (2s, 2H, OH); ¹³C NMR (75 MHz; DMSO- d_6) δ : 30.6 (N(1)CH₃), 101.7, 106.9, 110.9, 117.3, 125.2, 128.5, 128.7, 128.9, 132.0, 133.9, 158.9, 161.4 (Ph, CH=C), 160.9 (C=O), 174.1 (C=S). MS *m*/*z* 326 [M]⁺. HRMS (EI+) calcd for C₁₇H₁₄N₂O₃S [M]⁺: 326.0725; found 326.0724.

4.3.5. 3-Phenyl-5-(2,3-dihydroxybenzylidene)-1-methyl-2thiohydantoin (3by)

Use of thiohydantoin **11b** and aldehyde **8y** in the general procedure (80 °C, 7 h) afforded the title compound as a red solid: yield, 47 mg (71%); Mp 155–156 °C; R_f 0.18 (1:6:12 MeOH/EtOAc/hexanes); IR (KBr) 3355, 3062, 2938, 1732, 1595, 1471, 1350, 1283, 1233, 1154, 1052, 1004, 735, 529 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 3.35, 3.63 (2s, 3H, N(1)CH₃), 6.88, 7.04 (2s, 1H, CH=C), 6.56–6.87 (m, 2H, Ph), 7.32–7.73 (m, 6H, Ph), 9.07, 9.17 (2s, 1H, OH), 9.53, 9.65 (2s, 1H, OH); ¹³C NMR (75 MHz; DMSO- d_6) δ : 30.4, 33.8 (N(1)CH₃), 112.0, 116.1, 116.3, 116.7, 117.8, 118.7, 119.5, 119.6, 120.9, 121.0, 127.5, 127.6, 127.7, 128.4, 128.5, 128.6, 128.7, 133.5, 133.6, 144.3, 144.7, 145.1, 145.2 (Ph, CH=C, one peak was not detected and believed to overlap with the observed peaks), 160.6, 162.6 (C=O), 175.4, 179.0 (C=S). MS m/z 326 [M]⁺. HRMS (EI+) calcd for C₁₇H₁₄N₂O₃S [M]⁺: 326.0725; found 326.0722.

4.3.6. 3-Phenyl-5-(4-(*N*,*N*-dimethylamino)benzylidene)-1methyl-2-thiohydantoin (3bz)

Use of thiohydantoin **11b** and aldehyde **8z** in the general procedure (65 °C, 4 h) afforded the title compound as a red solid: yield, 61 mg (92%); Mp 238–240 °C; R_f 0.36 (1:2 EtOAc/hexanes); IR (KBr) 3748, 2368, 1715, 1581, 1530, 1501, 1478, 1393, 1344, 1194, 1147, 1056, 819, 763, 518 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 3.00 (s, 6H, N(CH₃)₂), 3.63 (s, 3H, N(1)CH₃), 6.72 (d, *J* = 9.0 Hz, 2H, Ph), 6.89 (s, 1H, CH=C), 7.36 (d, *J* = 7.8 Hz, 2H, Ph), 7.42–7.54 (m, 3H, Ph), 8.18 (d, *J* = 9.0 Hz, 2H, Ph); ¹³C NMR (75 MHz; DMSO- d_6) δ : 30.8 (N(1)CH₃), 39.6 (N(CH₃)₂), 111.0, 119.9, 124.2, 124.4, 128.5, 128.7, 128.9, 133.4, 134.0, 151.4 (Ph, CH=C), 160.9 (C=O), 173.4 (C=S). MS *m*/*z* 337 [M]⁺. HRMS (EI+) calcd for C₁₉H₁₉N₃OS [M]⁺: 337.1248; found 337.1248.

4.3.7. 3-(4-Methylphenyl)-5-(2,4-dihydroxybenzylidene)-1methyl-2-thiohydantoin (3cx)

Use of thiohydantoin **11c** and aldehyde **8x** in the general procedure (80 °C, 8 h) afforded the title compound as a red solid: yield, 44 mg (65%); Mp 275–279 °C; R_f 0.36 (1:1 EtOAc/hexanes); IR (KBr) 3748, 3444, 2917, 2381, 2324, 1688, 1593, 1511, 1463, 1398, 1350, 1275, 1175, 1105, 1002, 977, 947, 873, 802, 765, 677, 629, 588, 535, 499 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 2.36 (s, 3H, PhCH₃), 3.62 (s, 3H, N(1)CH₃), 6.24 (dd, *J* = 8.7, 2.4 Hz, 1H, Ph), 6.39 (d, *J* = 2.4 Hz, 1H, Ph), 7.05 (s, 1H, CH=C), 7.22 (d, *J* = 8.4 Hz, 2H, Ph), 7.30 (d, *J* = 8.4 Hz, 2H, Ph), 8.42 (d, *J* = 8.7 Hz, 1H, Ph), 10.00 (s, 1H, OH), 10.26 (s, 1H, OH); ¹³C NMR (75 MHz; DMSO- d_6) δ : 20.8 (PhCH₃), 30.6 (N(1)CH₃), 101.7, 106.9, 110.9, 117.2, 125.2, 128.6, 129.2, 131.3, 132.0, 138.1, 158.8, 161.4 (Ph, CH=C), 161.0 (C=O), 174.3 (C=S). MS *m*/*z* 340 [M]⁺. HRMS (EI+) calcd for C₁₈H₁₆N₂O₃S [M]⁺: 340.0881; found 340.0878.

4.3.8. 3-(4-Methylphenyl)-5-(2,3-dihydroxybenzylidene)-1methyl-2-thiohydantoin (3cy)

Use of thiohydantoin **11c** and aldehyde **8y** in the general procedure (80 °C, 9 h) afforded the title compound as a red solid: yield, 52 mg (78%); Mp 184–185 °C; R_f 0.60 (1:1 EtOAc/hexanes); IR (KBr) 3383, 3037, 2922, 2853, 2306, 2071, 1727, 1621, 1586, 1514, 1472, 1394, 1349, 1282, 1204, 1154, 1051, 1003, 942, 873, 764, 663, 523 cm⁻¹; ¹H NMR (300 MHz; CDCl₃) δ : 2.41, 2.43 (2s, 3H, PhCH₃), 3.41, 3.76 (2s, 3H, N(1)CH₃), 6.73–7.10 (m, 3H, Ph, CH=C), 7.15–7.41 (m, 5H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 21.5 (PhCH₃), 31.4, 34.2 (N(1)CH₃), 110.9, 116.4, 116.6, 118.7, 120.0, 120.5, 122.1, 122.5, 122.6, 124.8, 128.1, 128.3, 130.1, 130.2, 130.5, 130.8, 139.6, 140.1, 142.9, 143.3, 144.1, 148.5 (Ph, CH=C, two peaks were not detected and believed to overlap with the observed peaks), 163.7, 164.8 (C=O), 175.9, 180.2 (C=S). MS *m*/*z* 340 [M]⁺. HRMS (El+) calcd for C₁₈H₁₆N₂O₃S [M]⁺: 340.0881; found 340.0878.

4.3.9. 3-(4-Methylphenyl)-5-(4-(*N*,*N*-dimethylamino) benzylidene)-1-methyl-2-thiohydantoin (3cz)

Use of thiohydantoin **11c** and aldehyde **8z** in the general procedure (65 °C, 11 h) afforded the title compound as a red solid: yield, 46 mg (66%); Mp 241–243 °C; R_f 0.83 (1:1 EtOAc/hexanes); IR (KBr) 3748, 2917, 2853, 2382, 2091, 1714, 1619, 1580, 1515, 1478, 1441, 1390, 1194, 1146, 1053, 952, 888, 819, 763, 662, 516 cm⁻¹; ¹H NMR (300 MHz; CDCl₃) δ : 2.42 (s, 3H, PhCH₃), 3.06 (s, 6H, N (CH₃)₂), 3.51, 3.57 (2s, 3H, N(1)CH₃), 6.56, 7.03 (2s, 1H, CH=C), 6.65–6.77 (m, 3H, Ph), 7.23–7.36 (m, 3H, Ph), 8.15 (d, *J* = 9.0 Hz, 2H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 21.5 (PhCH₃), 31.1, 35.9 (N (1)CH₃), 40.3 (N(CH₃)₂), 111.6, 111.7, 118.3, 119.6, 120.3, 124.0, 125.2, 126.7, 128.2, 128.4, 129.9, 130.0, 131.2, 132.0, 133.8, 139.1, 139.3, 151.0, 151.8 (Ph, CH=C, one peak was not detected and believed to overlap with the observed peaks), 161.7, 163.9 (C=O), 174.9, 179.7 (C=S). MS *m*/*z* 351 [M]⁺. HRMS (EI+) calcd for C₂₀H₂₁N₃OS [M]⁺: 351.1405; found 351.1407.

4.3.10. 3-(4-Ethylphenyl)-5-(2,4-dihydroxybenzylidene)-1methyl-2-thiohydantoin (3dx)

Use of thiohydantoin **11d** and aldehyde **8x** in the general procedure (80 °C, 4 h) afforded the title compound as a red solid: yield, 43 mg (63%); Mp 235–239 °C; R_f 0.45 (1:1 EtOAc/hexanes); IR (KBr) 3434, 1627, 1513, 1391, 1348, 1155, 1105, 1053, 673, 497 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 1.22 (t, J = 7.6 Hz, 3H, PhCH₂CH₃), 2.66 (q, J = 7.6 Hz, PhCH₂), 3.61 (s, 3H, N(1)CH₃), 6.24 (dd, J = 8.7, 2.4 Hz, 1H, Ph), 6.39 (d, J = 2.4 Hz, 1H, Ph), 7.05 (s, 1H, CH=C), 7.24 (d, J = 8.4 Hz, 2H, Ph), 7.33 (d, J = 8.4 Hz, 2H, Ph), 8.41 (d, J = 8.7 Hz, 1H, Ph), 10.10 (br s, 2H, OH); ¹³C NMR (75 MHz; DMSO- d_6) δ : 15.4 (PhCH₂CH₃), 27.8 (PhCH₂), 30.6 (N(1)CH₃), 101.7, 106.9, 110.9, 117.2, 125.2, 128.6, 129.1, 131.5, 132.0, 144.2, 158.9, 161.4 (Ph, CH=C), 161.1 (C=O), 174.3 (C=S). MS m/z 354 [M]⁺. HRMS (EI+) calcd for C₁₉H₁₈N₂O₃S [M]⁺: 354.1038; found 354.1035.

4.3.11. 3-(4-Ethylphenyl)-5-(2,3-dihydroxybenzylidene)-1methyl-2-thiohydantoin (3dy)

Use of thiohydantoin **11d**, aldehyde **8y**, and additional AlCl₃ (0.040 mmol) and piperidine (0.60 mmol) in the general procedure (80 °C, 4 h and additionally, 4 h) afforded the title compound as a red solid: yield, 38 mg (53%); Mp 163–168 °C; R_f 0.51 (1:1 EtOAc/hexanes); IR (KBr) 3368, 2964, 1721, 1648, 1586, 1514, 1470, 1398, 1349, 1282, 1169, 1051, 1002, 939, 827, 763, 731, 692, 521 cm⁻¹; ¹H NMR (300 MHz; CDCl₃) δ : 1.14–1.23 (m, 3H, PhCH₂-CH₃), 2.53–2.71 (m, 2H, PhCH₂), 3.31, 3.64 (2s, 3H, N(1)CH₃), 6.62–7.03 (m, 4H, Ph, CH=C), 7.12–7.32 (m, 4H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 15.2 (PhCH₂CH₃), 28.7 (PhCH₂), 31.3, 34.3 (N(1)CH₃), 111.4, 116.3, 116.6, 118.5, 119.8, 120.4, 121.9, 122.2, 122.3,

124.6, 128.1, 128.2, 128.8, 128.9, 130.2, 130.3, 130.8, 143.0, 143.3, 144.2, 145.6, 146.1, 148.1 (Ph, CH=C, one peak was not detected and believed to overlap with the observed peaks), 163.9, 164.6 (C=O), 175.9, 180.0 (C=S). MS m/z 354 [M]⁺. HRMS (EI+) calcd for C₁₉H₁₈N₂O₃S [M]⁺: 354.1038; found 354.1041.

4.3.12. 3-(4-Ethylphenyl)-5-(4-(*N*,*N*-dimethylamino) benzylidene)-1-methyl-2-thiohydantoin (3dz)

Use of thiohydantoin **11d** and aldehyde **8z** in the general procedure (65 °C, 11 h) afforded the title compound as a red solid: yield, 145 mg (95%); Mp 160–162 °C; R_f 0.66 (1:1 EtOAc/hexanes); IR (KBr) 3435, 1710, 1621, 1581, 1514, 1477, 1388, 1344, 1193, 1145, 1052, 951, 887, 817, 515 cm⁻¹; ¹H NMR (300 MHz; CDCl₃) δ : 1.27 (t, J = 7.6 Hz, 3H, PhCH₂CH₃), 2.71 (q, J = 7.6 Hz, 2H, PhCH₂), 3.03 (s, 6H, N(CH₃)₂), 3.68 (s, 3H, N(1)CH₃), 6.52 (s, 1H, CH=C), 6.65 (d, J = 9.0 Hz, 2H, Ph), 7.23–7.36 (m, 4H, Ph), 8.14 (d, J = 9.0 Hz, 2H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 15.3 (PhCH₂CH₃), 28.7 (PhCH₂), 30.1 (N(1)CH₃), 40.2 (N(CH₃)₂), 111.6, 120.2, 124.0, 125.1, 128.4, 128.7, 131.3, 133.7, 145.2, 151.8 (Ph, CH=C), 161.7 (C=O), 174.8 (C=S). MS m/z 365 [M]⁺. HRMS (EI+) calcd for C₂₁H₂₃N₃OS [M]⁺: 365.1561; found 365.1564.

4.3.13. 3-(3-Chlorophenyl)-5-(2,4-dihydroxybenzylidene)-1methyl-2-thiohydantoin (3ex)

Use of thiohydantoin **11e** and aldehyde **8x** in the general procedure (80 °C, 5 h) afforded the title compound as a red solid: yield, 51 mg (70%); Mp 225–228 °C; R_f 0.17 (1:2 EtOAc/hexanes); IR (KBr) 3748, 3610, 2916, 2382, 2307, 1832, 1700, 1593, 1481, 1390, 1275, 1163, 975, 852, 748, 617, 501 cm⁻¹; ¹H NMR (300 MHz; DMSO-*d*₆) δ : 3.62 (s, 3H, N(1)CH₃), 6.25 (dd, *J* = 8.7, 1.8 Hz, 1H, Ph), 6.38 (d, *J* = 1.8 Hz, 1H, Ph), 7.06 (s, 1H, CH=C), 7.30–7.62 (m, 4H, Ph), 8.41 (d, *J* = 8.7 Hz, 1H, Ph), 10.03 (s, 1H, OH), 10.29 (s, 1H, OH); ¹³C NMR (75 MHz; DMSO-*d*₆) δ : 30.6 (N(1)CH₃), 101.7, 107.0, 110.8, 117.6, 125.1, 127.9, 128.7, 129.0, 130.2, 132.1, 132.7, 135.2, 158.9, 161.5 (Ph, CH=C), 160.6 (C=O), 173.7 (C=S). MS *m/z* 360 [M]⁺. HRMS (EI+) calcd for C₁₇H₁₃ClN₂O₃S [M]⁺: 360.0335; found 360.0336.

4.3.14. 3-(3-Chlorophenyl)-5-(2,3-dihydroxybenzylidene)-1methyl-2-thiohydantoin (3ey)

Use of thiohydantoin **11e** and aldehyde **8y** in the general procedure (80 °C, 3 h) with work-up and column chromatography afforded the title compound as a red solid: yield, 47 mg (65%); Mp 186–188 °C; R_f 0.23 (1:2 EtOAc/hexanes); IR (KBr) 3433, 1637, 1478, 1392, 1350, 1284, 1159, 1053 cm⁻¹; ¹H NMR (300 MHz; CDCl₃) δ : 3.42, 3.76 (2s, 3H, N(1)CH₃), 6.77–7.09 (m, 4H, Ph, CH=C), 7.26–7.48 (m, 4H, Ph); ¹³C NMR (75 MHz; CDCl₃) δ : 31.2, 34.1 (N(1)CH₃), 111.5, 116.2, 116.6, 118.6, 119.5, 120.2, 121.8, 121.9, 122.2, 124.4, 126.6, 126.7, 127.8, 128.6, 129.4, 129.8, 129.9, 130.0, 130.1, 133.5, 134.3, 134.6, 134.7, 143.0, 143.2, 143.8, 147.8, 161.1 (Ph, CH=C), 162.9, 163.9 (C=O), 174.9, 179.1 (C=S). MS *m/z* 360 [M]⁺. HRMS (EI+) calcd for C₁₇H₁₃ClN₂O₃S [M]⁺: 360.0335; found 360.0333.

4.3.15. 3-(3-Chlorophenyl)-5-(4-(*N*,*N*-dimethylamino) benzylidene)-1-methyl-2-thiohydantoin (3ez)

Use of thiohydantoin **11e** and aldehyde **8z** in the general procedure (65 °C, 10 h) with work-up and column chromatography afforded the title compound as a red solid: yield, 61 mg (79%); Mp 148–150 °C; R_f 0.39 (1:2 EtOAc /hexanes); IR (KBr) 3435, 1725, 1631, 1594, 1480, 1386, 1370, 1193, 1154, 1052, 950, 823, 517 cm⁻¹; ¹H NMR (300 MHz; DMSO-*d*₆) δ : 3.01 (s, 6H, N(CH₃)₂), 3.64 (s, 3H, N(1)CH₃), 6.75 (d, *J* = 9.0 Hz, 2H, Ph), 6.91 (s, 1H, CH=C), 7.32–7.45 (m, 1H, Ph), 7.50–7.61 (m, 3H, Ph), 8.17 (d, *J* = 9.0 Hz, 2H, Ph); ¹³C NMR (75 MHz; DMSO-*d*₆) δ : 30.6 (N(1)CH₃), 111.0, 119.6, 124.1, 124.3, 127.8, 128.5, 129.0, 130.1, 132.5, 133.3, 135.2, 151.3

(Ph, CH=C), 160.4, (C=O), 172.8 (C=S). MS *m*/*z* 371 [M]⁺. HRMS (EI+) calcd for C₁₉H₁₈ClN₃OS [M]⁺: 371.0859; found 371.0857.

4.3.16. 3-(Indan-5-yl)-5-(2,4-dihydroxybenzylidene)-1-methyl-2-thiohydantoin (3fx)

Use of thiohydantoin **11f** and aldehyde **8x** in the general procedure (80 °C, 1 h) afforded the title compound as a yellow solid: yield, 47 mg (63%); Mp 228–229 °C; R_f 0.29 (1:2 EtOAc/hexanes); IR (KBr) 3735, 3434, 1633, 1396, 1348, 704, 611, 518 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 1.96–2.12 (m, 2H, CH₂-indanyl), 2.82–2.94 (m, 4H, CH₂-indanyl), 3.60 (s, 3H, N(1)CH₃), 6.23 (dd, J = 9.0, 2.1 Hz, 1H, Ph), 6.36 (d, J = 2.1 Hz, 1H, Ph), 7.03 (s, 1H, CH=C), 7.04 (d, J = 7.8 Hz, 1H, Ph-indanyl), 7.15 (s, 1H, Ph-indanyl), 7.31 (d, J = 7.8 Hz, 1H, Ph-indanyl), 8.38 (d, J = 9.0 Hz, 1H, Ph); ¹³C NMR (75 MHz; DMSO- d_6) δ : 25.4 (CH₂-indanyl), 30.8 (N(1)CH₃), 32.2, 32.4 (CH₂-indanyl), 101.9, 107.1, 111.1, 117.3, 124.4, 124.8, 125.4, 126.8, 132.0, 132.2, 144.5, 159.0, 161.5 (Ph, Ph-indanyl, CH=C, one peak was not detected and believed to overlap with the observed peaks), 161.3 (C=O), 174.6 (C=S). MS m/z 366 [M]⁺. HRMS (EI+) calcd for C₂₀H₁₈N₂O₃S [M]⁺: 366.1038; found 366.1035.

4.3.17. 3-(Indan-5-yl)-5-(2,3-dihydroxybenzylidene)-1-methyl-2-thiohydantoin (3fy)

Use of thiohydantoin **11f** and aldehyde **8y** in the general procedure (80 °C, 5 h) afforded the title compound as a yellow solid: yield, 51 mg (83%); Mp 110–113 °C; R_f 0.45 (1:1 EtOAc/hexanes); IR (KBr) 3433, 2102, 1637, 1384, 1051, 531 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 1.98–2.13 (m, 2H, CH₂-indanyl), 2.82–2.95 (m, 4H, CH₂-indanyl), 3.33, 3.61 (2s, 3H, N(1)CH₃), 6.55–6.83 (m, 2H, Ph), 6.85, 7.01 (2s, 1H, CH=C), 7.02–7.67 (m, 4H, Ph, Ph-indanyl); ¹³C NMR (75 MHz; DMSO- d_6) δ : 25.4 (CH₂-indanyl), 30.8, 34.2 (N(1)CH₃), 32.2, 32.4 (CH₂-indanyl), 112.3, 116.4, 116.5, 117.0, 118.2, 119.1, 119.9, 120.0, 121.3, 121.4, 124.5, 124.6, 124.7, 124.8, 126.6, 126.8, 128.1, 129.1, 131.9, 132.0, 144.5, 144.6, 144.7, 144.8, 144.9, 145.1, 145.4, 145.5 (Ph, Ph-indanyl, CH=C), 161.1, 163.2 (C=O), 176.0, 179.7 (C=S). MS m/z 366 [M]⁺. HRMS (EI+) calcd for C₂₀H₁₈N₂O₃S [M]⁺: 366.1038; found 366.1035.

4.3.18. 3-(Indan-5-yl)-5-(4-(*N*,*N*-dimethylamino)benzylidene)-1-methyl-2-thiohydantoin (3fz)²³

Use of thiohydantoin **11f** and aldehyde **8z** in the general procedure (65 °C, 8 h) afforded the title compound as a yellow solid: yield, 46 mg (61%); Mp 129–131 °C; R_f 0.67 (1:1 EtOAc /hexanes); IR (KBr) 3455, 1715, 1635, 1599, 1489, 1387, 1353, 1159, 1122, 1050, 951, 822, 512 cm⁻¹; ¹H NMR (300 MHz; DMSO- d_6) δ : 1.98–2.13 (m, 2H, CH₂-indanyl), 2.80–2.95 (m, 4H, CH₂-indanyl), 3.00 (s, 6H, N(CH₃)₂), 3.63 (s, 3H, N(1)CH₃), 6.72 (d, *J* = 9.0 Hz, 2H, Ph), 6.88 (s, 1H, CH=C), 7.05 (d, *J* = 7.9 Hz, 1H, Ph-indanyl), 7.16 (s, 1H, Ph-indanyl), 7.32 (d, *J* = 7.9 Hz, 1H, Ph-indanyl), 8.17 (d, *J* = 9.0 Hz, 2H, Ph); ¹³C NMR (75 MHz; DMSO- d_6) δ : 25.1 (CH₂-indanyl), 30.6 (N(1)CH₃), 31.9, 32.1 (CH₂-indanyl), 111.0, 119.8, 123.9, 124.1, 124.3, 124.5, 126.5, 131.9, 133.2, 144.2, 151.2 (Ph, Ph-indanyl, CH=C, one peak was not detected and believed to overlap with the observed peaks), 160.9 (C=O), 173.6 (C=S). MS *m/z* 377 [M]⁺. HRMS (El+) calcd for C₂₂H₂₃N₃OS [M]⁺: 377.1561; found 377.1565.

4.4. General procedure for evaluation of NOX 1 and 4 inhibitory activities

We established transgenic fly lines overexpressing human NOX 1 or human NOX 4 in a *Drosophila* DUOX (dDUOX) knockdown condition according to the previous procedure.³³ The genotypes of lines used in this study were human NOX 1 (*UAS-hNOX1/UAS-dDUOX-RNAi; Da-GAL4/+*) and human NOX 4 transgenic fly (*UAS-hNOX4/UAS-dDUOX-RNAi; Da-GAL4/+*). We detected human NOX 1 or human NOX 4 isozyme in each transgenic fly using real-

time quantitative PCR (qPCR). Transgenic flies were homogenated with ice-cold PBS containing protease inhibitors and membranes enriched human NOX 1 or NOX 4 were harvested. Membranes containing human NOX 1 or NOX 4 served to monitor ROS production by lucigenin chemiluminescence in the absence or presence of chemical compound. The reaction medium consisted in Hepes-buffered salt solution (128 mM NaCl, 2.5 mM KCl, 1.2 mM MgSO₄, 1.0 mM KH₂PO₄, 1.75 μ M CaCl₂, 0.03 Na₂EDTA, 5.5 mM glucose, and 20 mM HEPES, pH 7.4), 400 μ M lucigenin (10,10-dimethylbis-9,9-bisacridinium nitrate) and 500 μ M NADPH.³⁴

Acknowledgements

This work was supported by the National Research Foundation of Korea (NRF) grants (2012R1A5A1048236, 2012R1A1A2001033 and 2011-0028885), by Redoxomics grant (2012M3A9C5048708), and by Priority Research Centers Program (2016R1A6A1A03007648) through NRF funded by the Ministry of Education, Science and Technology (MEST) – Republic of Korea.

References and notes

- 1. Kiec-Kononowicz, C.; Szymanska, E. Farmaco 2002, 57, 909.
- Khodair, A. I.; El-Barbary, A. A.; Abbas, Y. A.; Imam, D. R. Phosphorus Sulfur Silicon Relat. Elem. 2001, 170, 261.
- 3. Mehta, N.; Risinger, C. A.; Soroko, F. E. J. Med. Chem. 1981, 24, 465.
- 4. Struck, R. F.; Kirk, M. C.; Rice, L. S.; Suling, W. J. J. Med. Chem. 1986, 29, 1319.
- Cave, A. C.; Brewer, A. C.; Narayanapanicker, A.; Ray, R.; Grieve, D. J.; Walker, S.; Shah, A. M. Antioxid. Redox Signal. 2006, 8, 691.
- 6. Bedard, K.; Krause, K.-H. Physiol. Rev. 2007, 87, 245.
- VanBuul, J. D.; Fernandez-Borja, M.; Anthony, E. C.; Hordijk, P. L. Antioxid. Redox Signal. 2005, 7, 308.
- Guzik, T. J.; Sadowski, J.; Guzik, B.; Jopek, A.; Kapelak, B.; Przybylowski, P.; Wierzbicki, K.; Korbut, R.; Harrison, D. G.; Channon, K. M. Arterioscler. Thromb. Vasc. Biol. 2006, 26, 333.
- Jaquet, V.; Scapozza, L.; Clark, R.; Krause, K. H.; Lambeth, J. D. Antioxid. Redox Signal. 2009, 11, 2535.
- Borbely, G.; Szabadkai, I.; Horvath, Z.; Marko, P.; Varga, Z.; Breza, N.; Baska, F.; Vantus, T.; Huszar, M.; Geiszt, M.; Hunyady, L.; Buday, L.; Orfi, L.; Keri, G. *J. Med. Chem.* **2010**, *53*, 6758.

- Freyhaus, H.; Huntgeburth, M.; Wingler, K.; Schnitker, J.; Baumer, A. T.; Vantler, M.; Bekhite, M. M.; Wartenberg, M.; Seuer, H.; Rosenkranz, S. *Cardiovasc. Res.* 2006, 71, 331.
- 12. Stielow, C.; Catar, R. A.; Muller, G.; Wingler, K.; Scheurer, P.; Schmidt, H. H.; Morawietz, H. *Biochem. Biophys. Res. Commun.* **2006**, 344, 200.
- Ciolkowski, M.; Paneth, P.; Lorenz, I.-P.; Mayer, P.; Rozalski, M.; Krajewska, U.; Budzisz, E. J. Enzym. Inhib. Med. Chem. 2009, 24, 1257.
- 14. Bin, H.-Y.; Bae, Y. S.; Kim, G.; Lee, K.-I. Synthesis 2011, 11, 1783.
- Joo, J. H.; Huh, J.-E.; Lee, J. H.; Park, D. R.; Lee, Y.; Lee, S. G.; Choi, S.; Lee, H. J.; Song, S.-W.; Jeong, Y.; Goo, J.-I.; Choi, Y.; Baek, H. K.; Yi, S. S.; Park, S. J.; Lee, J. E.; Ku, S. K.; Lee, W. J.; Lee, K.-I.; Lee, S. Y.; Bae, Y. S. Sci. Rep. 2016, 6, 22389.
- Bae, Y. S.; Lee, K.-I.; Lee, S. Y.; Choi, S.; Lee, J. H. PCT Int. Appl. 2009, WO 2009119987 A2 20091001.
- 17. Rowley, G. L.; Greenleaf, A. L.; Kenyon, G. J. J. Am. Chem. Soc. 1971, 93, 5542.
- 18. Boulos, L. S.; Yakout, E.-S. M. A.; Arsanious, M. H. N. Phosphorus Sulfur Silicon Relat. Elem. 2006, 181, 1615.
- Silva, T. G.; Barbosa, F. S. V.; Brandao, S. S. F.; Lima, M. C. A.; Galdino, S. L.; Pitta, I. R.; Barbe, J. Heterocycl. Commun. 2001, 7, 523.
- Aly, Y. L.; El-Shehawy, A. A.; Attia, A. M.; Sobh, M. E. Phosphorus Sulfur Silicon Relat. Elem. 2011, 186, 1572.
- El-Regal, M. K. A.; Abdalha, A. A.; El-Kassaby, M. A.; Ali, A. T. Phosphorus Sulfur Silicon Relat. Elem. 2007, 182, 845.
- Elokdah, H.; Sulkowski, T. S.; Abou-Gharbia, M.; Butera, J. A.; Chai, S.-Y.; McFarlane, G. R.; McKean, M.-L.; Babiak, J. L.; Adelman, S. J.; Quinet, E. M. J. Med. Chem. 2004, 47, 681.
- Gregg, B. T.; Earley, W. G.; Golden, K. C.; Quinn, J. F.; Razzano, D. A.; Rennells, W. M. Synthesis 2006, 4200.
- 24. Zhang, X.; Lee, Y. K.; Kelley, J. A.; Burke, T. R., Jr. J. Org. Chem. 2000, 65, 6237.
- 25. Jeffreys, R. A. J. Chem. Soc. 1954, 389.
- 26. Cook, A. H.; Cox, S. F. J. Chem. Soc. 1949, 2342.
- 27. Jeffreys, R. A. J. Chem. Soc. 1954, 2221.
- The compound was known as one member of chemical library: Aurora Screening Library, Aurora Fine Chemicals LLC, San Diego, CA, USA, http://www. aurorafinechemicals.com.
- 29. Khodair, A. I.; Nielsen, J. Heterocycles 2002, 57, 1017.
- Gregg, B. T.; Golden, K. C.; Quinn, J. F.; Tymoshenko, D. O.; Earley, W. G.; Maynard, D. A.; Razzano, D. A.; Rennells, W. M.; Butcher, J. J. Comb. Chem. 2007, 9, 1036.
- Bourahla, K.; Paquin, A.; Lozach, O.; Meijer, L.; Carreaux, F.; Bazureau, J. P. Molecules 2011, 16, 7377.
- 32. The compounds were known as members of chemical library: AMRI Smartsourcing[™], AMRI Co., New York, NY, USA, http://www.amriglobal.com.
- **33.** Ha, E. M.; Oh, C. T.; Bae, Y. S.; Lee, W. J. *Science* **2005**, *310*, 847.
- 34. Rhee, S. G.; Chang, T. S.; Jeong, W.; Kang, D. Mol. Cells 2010, 29, 539.